

This Page Is Inserted by IFW Operations
and is not a part of the Official Record

BEST AVAILABLE IMAGES

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images may include (but are not limited to):

- BLACK BORDERS
- TEXT CUT OFF AT TOP, BOTTOM OR SIDES
- FADED TEXT
- ILLEGIBLE TEXT
- SKEWED/SLANTED IMAGES
- COLORED PHOTOS
- BLACK OR VERY BLACK AND WHITE DARK PHOTOS
- GRAY SCALE DOCUMENTS

IMAGES ARE BEST AVAILABLE COPY.

**As rescanning documents *will not* correct images,
please do not report the images to the
Image Problem Mailbox.**

(19)



Europäisches Patentamt
European Patent Office
Office européen des brevets



(11)

EP 1 127 578 A1

(12)

EUROPEAN PATENT APPLICATION
published in accordance with Art. 158(3) EPC

(43) Date of publication:

29.08.2001 Bulletin 2001/35

(51) Int Cl.⁷: **A61K 38/19, A61K 47/36,**
A61P 19/08, A61P 3/00
// C07K14:52

(21) Application number: **99952793.0**

(22) Date of filing: **28.10.1999**

(86) International application number:
PCT/JP99/05963

(87) International publication number:
WO 00/24416 (04.05.2000 Gazette 2000/18)

(84) Designated Contracting States:

AT BE CH CY DE DK ES FI FR GB GR IE IT LI LU
MC NL PT SE

Designated Extension States:
AL LT LV MK RO SI

(30) Priority: **28.10.1998 JP 32287498**

(71) Applicants:

- **SNOW BRAND MILK PRODUCTS, CO., LTD.**
Sapporo-shi Hokkaido 065-0043 (JP)
- **Sankyo Co., Ltd.**
Tokyo 140-8710 (JP)

(72) Inventors:

- **MOCHIZUKI, Shinichi**
Kawachi-gun, Tochigi 329-0433 (JP)

- **FUJISE, Nobuaki, Ishibashi Heights 201**
Shimotsuga-gun, Tochigi 329-0511 (JP)
- **MASUYAMA, Chiharu**
Shimotsuga-gun, Tochigi 329-0511 (JP)
- **TSUDA, Eisuke,**
Daiyaparesu Jichiidai 5-bankan 407
Kawachi-gun, Tochigi 329-0434 (JP)
- **HIGASHIO, Kanji**
Kawagoe-shi, Saitama 350-0822 (JP)

(74) Representative: **Wakerley, Helen Rachael**
Reddie & Grose,
16 Theobalds Road
London WC1X 8PL (GB)

(54) **REMEDIES FOR BONE METABOLIC ERRORS**

(57) A novel bone-pathobolism treating agent is provided. A bone-pathobolism treating agent comprising at least one substance selected from the group consisting of osteoclastogenesis inhibitory factor (OCIF), its homologs, and its variants and a polysaccharide or its derivatives. As the polysaccharide or its derivatives,

heparin, dextran sulfate and the like can be used.

A bone-pathobolism treating agent is provided which has excellent therapeutic effect on bone-pathobolism such as osteoporosis, hypercalcemia, or chronic articular rheumatism and persistence of the activity. The agent is useful as a medicine.

Description

TECHNICAL FIELD

[0001] The present invention relates to a novel bone-pathobolism treating agent having high activity and high persistence. The bone-pathobolism treating agent of the present invention has excellent therapeutic activity on bone-pathobolism such as osteoporosis, hypercalcemia, or chronic articular rheumatism and is useful as a medicine.

BACKGROUND ART

[0002] Bones not only have an ability of supporting the body but also function as the largest storage-organ of the calcium in the organism and 99% of the calcium present in the organism is accumulated in the bones. In addition, bones are always remodelled through opposite actions of bone resorption and bone formation. This plays an important role on the maintenance of homeostasis of serum calcium. It is known that the activation of osteoclasts which bear an important role in bone resorption causes excessive flow of calcium into blood from bones to break the homeostasis of calcium in blood and induces hypercalcemia. Hypercalcemia is a disease which occurs due to osteectopia of tumor and the number of patients who suffer from it is expected to increase so that development of a treating agent is desired to be created in a hurry. At present, calcitonin, its derivatives, and bisphosphonate derivatives are used as such hypercalcemia treating agents. However, their therapeutic effect is not satisfactory and development of novel drugs replacing them is desired.

[0003] On the other hand, it has been reported that osteoclastogenesis inhibitory factor (OCIF) (WO96/26217) known as a protein inhibiting differentiation of osteoclasts has a hypocalcemic action [Biochemical and Biophysical Research Communications, Vol. 245, pp382-387 (1998); Endocrinology, Vol. 139, pp4012-4015 (1998)]. OCIF is expected as a quite new agent treating hypercalcemia. However, since it is a protein, OCIF is metabolized rapidly in the organism. Accordingly, development of an OCIF-preparation which is safer and has more enhanced action has been desired.

DISCLOSURE OF THE INVENTION

[0004] Under the circumstances, the inventors of the present invention have made extensive investigation and as a result, they have found that the effect of OCIF on bone-pathobolism can be further increased by adding a polysaccharide to OCIF to form a preparation. Therefore, an object of the present invention is to provide a bone-pathobolism treating agent in which the effect of OCIF on bone-pathobolism has been further increased and the effect has been rendered persistent.

[0005] The present invention relates to a bone-

pathobolism treating agent comprising both at least one substance selected from the group consisting of osteoclastogenesis inhibitory factor (OCIF), its homologs, and its variants, and a polysaccharide or its derivatives.

[0006] In the present invention, heparin is preferred as the above polysaccharide, and dextran sulfate is preferred as the above polysaccharide derivative.

[0007] According to the present invention, there is provided a therapeutic agent having excellent action on bone-pathobolism such as osteoporosis, hypercalcemia, or chronic articular rheumatism and persistence of the activity. The therapeutic agent is useful as a medicine.

[0008] The present invention relates to a method for increasing the activity of osteoclastogenesis inhibitory factor using a polysaccharide or its derivatives.

[0009] OCIF used in the present invention is natural type or recombinant type obtained by the method described in WO96/26217 and its origin is not particularly limited. But particularly preferred OCIF is human type OCIF. Such a natural or recombinant type OCIF includes a monomer type having a molecular weight of about 60 kDa and a dimer type having a molecular weight of about 120 kDa as measured by SDS-PAGE under non-reducing conditions.

[0010] In the present invention, analogs and variants of OCIF may be used. The homologs include those obtained by preparing a cDNA library using poly (A)⁺ RNA from IMR-90 cell (ATCC CCL-186), obtaining cDNA of OCIF homolog by a hybridization method using OCIF cDNA fragment as a probe, inserting the cDNA into an expression vector, introducing the vector in a usually used host, expressing the cDNA in the host, and purifying the expressed protein by a conventional method. More specifically, the homologs include OCIF2, OCIF3, OCIF4, or OCIF5 as described in WO96/26217.

[0011] As described in WO96/26217, among these, OCIF2 has a deletion of 21 bp from the 265th guanine to the 285th guanine in the base sequence of OCIF cDNA and a deletion of 7 amino acids from the 68th glutamic acid (Glu) to the 74th glutamine (Gln) in the amino acid sequence.

[0012] OCIF3 has the same base sequence as the OCIF cDNA except for the followings. The 9th cytidine is replaced by guanine and the -19th asparagine (Asn) is replaced by lysine (Lys) in the amino acid sequence. However, this is the substitution of amino acids in the signal sequence and is considered to have no influence on OCIF3 to be secreted. Further, OCIF3 has a deletion of 117 bp from the 872nd guanine to the 988th guanine in the base sequence of OCIF cDNA and a deletion of 39 amino acids from the 270th threonine (Thr) to the 308th leucine (Leu) in the amino acid sequence.

[0013] OCIF4 has the same base sequence as OCIF cDNA except for the followings. The 9th citidine is replaced by guanine and the -19th asparagine (Asn) is replaced by lysine (Lys) in the amino acid sequence. The 22nd guanine is replaced by thymidine and the -14th

alanine (Ala) is replaced by serine (Ser) in the amino acid sequence of OCIF. However, this is the substitution of amino acids in the signal sequence and is considered to have no influence on OCIF4 to be secreted. Further, there is an insertion of intron 2 of about 4 kb between the 400th and 401th in the base sequence of OCIF cDNA and the open reading frame is terminated therein. In the amino acid sequence, a novel amino acid sequence consisting of 21 amino acids is added after the 112th alanine (Ala) of the amino acid sequence of OCIF.

[0014] OCIF5 has the same base sequence as OCIF cDNA except for the followings. The 9th cytidine is replaced by guanine and the -19th asparagine (Asn) is replaced by lysine (Lys) in the amino acid sequence. However, this is the substitution of amino acids in the signal sequence and is considered to have no influence on OCIF5 to be secreted. Further, there is an insertion of the latter half of intron 2 of about 1.8 kb between the 400th and 401th in the base sequence of OCIF cDNA and the open reading frame is terminated therein. In the amino acid sequence, a novel amino acid sequence consisting of 12 amino acids is added after the 112th alanine (Ala) of the amino acid sequence of OCIF.

[0015] The variants include those in which one or more amino acids have been inserted in, added to, substituted in, or deleted of the amino acid sequence of OCIF. More specifically, the variants include those obtained by preparing OCIF variant cDNA by introducing substitution or deletion mutation by a PCR method or cleavage with a restriction enzyme, inserting the cDNA into an expression vector, incorporating the vector into a usually used host, expressing the cDNA in the host and purifying the expressed protein by a conventional method.

[0016] The polysaccharide used in the present invention is a polymer (glucan) formed through glycoside bonding of monosaccharides and is preferably a heteropolysaccharide (heteroglycan) having 2 or more constituent monosaccharides. More specifically, the polysaccharide which can be used include natural polysaccharides such as hyaluronic acid, chondroitin sulfate, dermatan sulfate, heparan sulfate, keratan sulfate, carrageenan, pectin, and heparin, synthetic polysaccharides such as dextran, and synthetic polysaccharide derivatives such as dextran sulfate. Particularly preferably, sulfate ester of glucan is used. For example, heparin having a molecular weight of 3,000 to 6,000 or dextran sulfate having a molecular weight of 5,000 to 10,000 is used. The bone-pathobolism treating agent of the present invention is preferably a combination of at least one substance selected from the group consisting of OCIF, its homologs, and its variants and a polysaccharide or its derivatives in a proportion of 1 to 100 folds, particularly 1 to 16 folds in an amount of the polysaccharide or its derivatives to the OCIF, its derivatives, or variants. The preparation of the present invention comprising a combination of at least one substance selected from the group consisting of OCIF, its analogs,

and its variants and a polysaccharide or its derivatives is a bone-pathobolism treating agent having excellent persistence and therapeutic effect as compared with administration of OCIF alone and is effective on bone-pathobolisms such as osteoporosis, hypercalcemia, chronic articular rheumatism.

[0017] The preparation of the present invention is safely administered orally or parenterally to humans or animals as a medicine. Parenteral administration includes intravenous injection, intramuscular injection, subcutaneous injection, nasal administration, intraoral administration, permucosubcutaneous administration, etc. The preparations to be administered by these administration routes can be formulated by a known pharmaceutical production method, and administered together with pharmacologically acceptable vehicles, excipients, lubricants, colorants, etc. as a preparation of a medical composition. When an injection is prepared, OCIF and a polysaccharide, optionally a pH adjuster, a buffer, a stabilizer, a solubilizer, etc. are added to form injections by a conventional method. In this case, known additives such as human serum albumin and a surfactant may be used in combination. As the surfactant, polyanions and anionic surfactants are cited. The injection can be dispensed in vials to form a solution preparation or prepared as a freeze-dried preparation which upon use is dissolved in distilled water, physiological saline or the like at appropriate timing. Upon administering OCIF to normal rats once a day for continuous 2 weeks in a dose of 3 or 24 mg/kg·day, increases in bone density and amount, respectively, were observed but no histopathological disorder in 38 tissues nor change in blood was observed [H. Yasuda et al.: Endocrinology, Vol. 139, pp1329-1337 (1998)]. Thus, the action of OCIF is highly specific to bone and it is expected that OCIF can be administered safely to humans.

[0018] The amount and method of administration of the bone-pathobolism treating agent of the present invention to patients are not limited particularly, since they may vary depending on the severity of symptom, age, condition of body, and body weight of the patient. For example, the agent may be parenterally administered once to several times a day in a dose of about 0.01 to 1 mg/kg per day for adults. The activity of the preparation of the present invention can be performed by measuring the concentration of serum calcium. For example, OCIF solution prepared with a suitable solvent and being added a polysaccharide is intravenously administered to a rat, blood is collected on time, and the serum calcium level is measured by a conventional method.

[0019] The present invention relates to a method for elevating the activity of osteoclastogenesis inhibitory factor by use of a polysaccharide or its derivatives. According to the present invention, the blood level of OCIF can be elevated to enhance the action of OCIF lowering the serum calcium level.

BRIEF DESCRIPTION OF THE DRAWINGS

[0020] Fig. 1 shows serum calcium level after 3 hours from the administration of the preparation containing OCIF and a polysaccharide in Example 2.

[Explanation of symbols]

[0021]

D-1: 0.5 mg/kg OCIF + 2 mg/kg dextran sulfate (molecular weight 5,000)

D-2: 0.5 mg/kg OCIF + 2 mg/kg dextran sulfate (molecular weight 8,000)

D-3: 0.5 mg/kg OCIF + 2 mg/kg dextran sulfate (molecular weight 10,000)

H-1: 0.5 mg/kg OCIF + 2 mg/kg heparin (207.8 units/mg)

H-2: 0.5 mg/kg OCIF + 2 mg/kg heparin (171.2 units/mg)

H-3: 0.5 mg/kg OCIF + 2 mg/kg heparin (molecular weight 3,000)

H-4: 0.5 mg/kg OCIF + 2 mg/kg heparin (molecular weight 6,000)

** : Significant (p values $\leq 1\%$)

[0022] Fig. 2 shows serum calcium level after 3 hours from the administration of OCIF and polysaccharide with various mixing ratios in Example 3.

[Explanation of symbols]

[0023] ** : Significant (p values $\leq 1\%$)

[0024] Fig. 3 shows serum calcium level after 3, 6, and 9 hours, respectively, from the administration of the preparation containing OCIF and a polysaccharide in Example 4.

[Explanation of symbols]

[0025] ** : Significant (p values $\leq 1\%$)

[0026] Fig. 4 shows time-dependent change in blood OCIF level when a preparation containing OCIF and a polysaccharide was administered in Example 5.

[Explanation of symbols]

[0027]

A : A figure illustrating the blood level of dimer type OCIF.

B : A figure illustrating the blood level of monomer type OCIF.

● : OCIF

○ : OCIF + dextran sulfate

[0028] Fig. 5 shows time-dependent change in the proportion of monomer type OCIF/dimer type OCIF in

blood when a preparation containing OCIF and a polysaccharide was administered in Example 5.

[Explanation of symbols]

[0029]

A : OCIF

B : OCIF + dextran sulfate

O : Dimer type OCIF

Δ : Monomer type OCIF

[0030] Fig. 6 shows blood OCIF level after 2 and 4 hours from the administration of a preparation containing OCIF and a polysaccharide (dextran sulfate, apple pectin, or citrus pectin) in Example 6.

[Explanation of symbols]

[0031]

□ : Administration of 0.5 mg/kg OCIF alone

◇ : 0.5 mg/kg OCIF + 0.5% dextran sulfate

○ : 0.5 mg/kg OCIF + 0.5% apple pectin

Δ : 0.5 mg/kg OCIF + 0.5% citrus pectin

[0032] Fig. 7 shows the blood OCIF level after 2 and 4 hours from the administration of a preparation containing OCIF and a polysaccharide (dextran sulfate, apple pectin, or carrageenan) in Example 6.

[Explanation of symbols]

[0033]

□ : Administration of 0.5 mg/kg OCIF alone

◇ : 0.5 mg/kg OCIF + 0.5% dextran sulfate

○ : 0.5 mg/kg OCIF + 0.5% apple pectin

Δ : 0.5 mg/kg OCIF + 0.5% carrageenan (λ)

[0034] Fig. 8 shows the blood OCIF level after intravenous administration of a preparation containing OCIF and a polysaccharide (dextran sulfate or apple pectin) in Example 7.

[Explanation of symbols]

[0035]

○ : Administration of 50 μ g/kg OCIF alone

Δ : 50 μ g/kg OCIF + 0.1% dextran sulfate

□ : 50 μ g/kg OCIF + 0.15% apple pectin

[0036] Fig. 9 shows the blood OCIF level after the intramuscular administration of a preparation containing OCIF and a polysaccharide (dextran sulfate or apple pectin)- in Example 7.

[Explanation of symbols]

[0037]

- : Administration of 1 mg/kg OCIF alone
 △: 1 mg/kg OCIF + 0.1% dextran sulfate
 □: 1 mg/kg OCIF + 0.15% apple pectin

[0038] Fig. 10 shows the serum calcium level after 4 hours from the administration of a preparation containing OCIF and a polysaccharide (apple pectin) in Example 8.

BEST MODE FOR CARRYING OUT THE INVENTION

[Examples]

[0039] The following examples are presented in order to more specifically explain the present invention by examples. However, they are only concrete illustrations and the present invention is by no means limited thereto.

[Example 1]

Production of Injections - 1

[0040] 500 µg of human OCIF obtained by the method described in WO96/26217 and 2 mg of heparin were dissolved in 5 ml of 10 mM sodium phosphate buffer solution (pH 7.0) containing 0.15 M NaCl and 0.01% Tween 80, and the resulting solution was sterilized by passing through a 0.22 µm sterile filter (Millex GV, Millipore Co.) and then packed in a vial to obtain injections for intravenous injection.

Production of Injections - 2

[0041] 500 µg of human OCIF obtained by the method described in WO96/26217 and 2 mg of dextran sulfate were dissolved in 5 ml of 10 mM sodium phosphate buffer solution (pH 7.0) containing 0.15 M NaCl and 0.01% Tween 80 and the resulting solution was sterilized by passing through a 0.22 µm sterile filter (Millex GV, Millipore Co.) and then packed in a vial to obtain injections for intravenous injection.

[Example 2]

Hypocalcemic Effect of OCIF by the Addition of a Polysaccharide - 1

[0042] To 2 ml of 0.25 mg/ml human OCIF solution prepared by dissolving human OCIF (of dimer type) (recombinant type OCIF obtained by the method described in WO96/26217) in 10 mM sodium phosphate buffer solution (pH 7.0) containing 0.15 M NaCl and 0.01% Tween 80 (hereafter, referred to as a solvent), 2 mg of dextran sulfate (molecular weight 8,000 or 10,000: Sig-

ma AB, and molecular weight 5,000 or 50,000: Wako Pure Chemical Industry Co., Ltd.), or heparin (207.8 or 171.2 unit/mg: Wako Pure Chemical Industry Co., Ltd. and molecular weight 3,000 or 6,000: Sigma AB) was dissolved and then the solution was incubated at 4°C for a day. At the same time, 0.25 and 2.5 mg/ml human type OCIF solutions and the solvent were incubated at 4°C for a day similarly. These test sample solutions were provided as OCIF + dextran sulfate administered group (D group), OCIF + heparin administered group (H group) (D group and H group were administered with 0.5 mg/kg OCIF), OCIF alone administered group (administered with 0.5 mg/kg and 5 mg/kg OCIF, respectively), and solvent administered group. Four weeks age female Wistar rats were once intravenously administered with the samples at a dose of 2 ml/kg. After 3 hours from the administration, blood was collected from the eyehole to prepare serum. The calcium level in the obtained serum was measured using Calcium C Test (Wako Pure Chemical Industry Co., Ltd.). Fig. 1 shows the results. As a result, it was observed that the administration of 0.5 mg/kg of human OCIF to which one of various types of dextran sulfate or heparin has been added exhibited a significant enhancing effect in hypocalcemic action. Therefore, it was confirmed that the addition of a polysaccharide can enhance hypocalcemic action of human OCIF.

[Example 3]

Effect of the amount of added polysaccharide on enhancement of the OCIF-action

[0043] In 2 ml of the 0.25 mg/ml human OCIF solutions prepared in the same manner as in Example 2, dextran sulfate (molecular weight 5,000: Wako Pure Chemical Industry Co., Ltd.) was dissolved in a proportion of 1, 2, 4, 8, or 16 folds in weight, based on the amount of OCIF respectively, and the mixtures were incubated at 4°C for a day. Similarly, heparin (207.8 units/mg: Wako Pure Chemical Industry Co., Ltd.) was dissolved in 2ml of 0.25 mg/ml human OCIF solutions in the same proportion and the mixtures were incubated at 4°C for a day.

[0044] Further, 4 mg/ml dextran sulfate or heparin solution, 0.25 and 2.5 mg/ml human OCIF solutions, and the solvent alone were similarly incubated at 4°C for a day. These test sample solutions were once intravenously administered to four weeks age female Wistar rats at a dose of 2 ml/kg. After 3 hours from the administration, blood was collected from the eyehole to prepare serum. The calcium level in the obtained serum was measured using Calcium C Test (Wako Pure Chemical Industry Co., Ltd.). The kit is constructed based on a chelate method (orthocresol phthalein coplexon (OCPC) method) in which the effect of magnesium is removed and specificity is increased with the addition of 8-quinolinol. Calcium give rises purple red color upon binding to OCPC under alkaline conditions. Measure-

ment of absorbance enables assessment of calcium concentration. Fig. 2 shows the results. As a result, significant hypocalcemic action was recognized when dextran sulfate was added 4 folds as much as human OCIF or more. Significant hypocalcemic action was also seen when an equivalent amount of heparin was added to human OCIF. Therefore, it was confirmed that simultaneous administration of human OCIF and a polysaccharide in specific proportions can further enhance the hypocalcemic action of OCIF.

[Example 4]

Enhancing effect on persistence of OCIF by the addition of a polysaccharide

[0045] After dissolving 20 mg of dextran sulfate (molecular weight 5,000: Wako Pure Chemical Industry Co., Ltd.) in 2 ml of 2.5 mg/ml human OCIF solution prepared with the solvent, the mixture was incubated at 4°C for a day. Twenty mg heparin (207.8 units/mg: Wako Pure Chemical Industry Co., Ltd.) was also dissolved in 2 ml of the 2.5 mg/ml human OCIF solution and similarly incubated at 4°C for a day. Further, 2.5 mg/ml human OCIF solution and the solvent were incubated at 4°C for a day. Four weeks age female Wistar rats were intravenously administered with the test sample solutions at a dose of 2 ml/kg (5 mg/kg as the amount of OCIF) respectively. After 3, 6, and 9 hours from the administration, blood was collected from the eyehole to prepare serum. The calcium level in the obtained serum was measured using Calcium C Test (Wako Pure Chemical Industry Co., Ltd.). Fig. 3 shows the results. As a result, a significant decrease in serum calcium level was observed in the group administered with 5 mg/kg human OCIF solution alone after 3 hours from the administration, but no significant hypocalcemic action was observed after 6 and 9 hours, respectively, after the administration.

[0046] On the other hand, 2.5 mg/ml human OCIF solution to which dextran sulfate or heparin was added, respectively, in amounts 4 folds as much as human OCIF had a significant hypocalcemic effect even after 9 hours from the administration. Therefore, it was confirmed that the simultaneous addition of human OCIF and a polysaccharide can give persistence enhancing effect.

[Example 5]

Enhancing effect of added polysaccharides on the persistence of circulating OCIF level-1

[0047] To 1 ml of the 1 mg/ml human OCIF solution prepared in the same manner as in Example 2, 1 ml of 4 mg/ml of dextran sulfate was added and the obtained mixture was incubated at 4°C for a day. The test sample solution was intravenously administered to 9 weeks age

male Wistar rats at a dose of 1 ml/kg. After 2, 5, 10, 15, 30, 45, 60, 120, 240, 360, 480, 600, 720, and 1440 minutes from the administration, blood was collected from the eyehole to prepare serum. The human OCIF level in the obtained serum was measured by the ELISA described in WO96/26217 using monoclonal antibodies capable of recognizing dimer type OCIF, monoclonal antibodies capable of recognizing monomer type OCIF (Biochemical and Biophysical Research Communications, Vol. 245, pp382-387 (1998)). The total OCIF level was calculated as the sum of the dimer type OCIF level and monomer type OCIF level. Figs. 4 and 5 show the results. As a result, as compared with the group administered with 500 µg/kg human OCIF alone, the group administered with the human OCIF solution to which dextran sulfate 4 folds as much as the human OCIF was added maintained evidently high circulating OCIF level (Fig. 4). It was confirmed that the conversion of dimer type OCIF to monomer type OCIF in blood was inhibited (Fig. 5). Therefore, the addition of a polysaccharide enables circulating OCIF level, in particular the circulating level of dimer type OCIF having high hypocalcemic activity (Biochemical and Biophysical Research Communications, Vol. 245, pp382-387 (1998)), to persist.

[Example 6]

Enhancing effect of added polysaccharides on the persistence of circulating OCIF level-2

[0048] To 2 ml of the 0.25 mg/ml human OCIF solution prepared in the same manner as in Example 2, equivalent volume of 0.5% solution [solvent: 10 mM sodium phosphate buffer solution (pH 7.0) containing 0.15 M NaCl, 0.01% polysorbate 80] of dextran sulfate, apple pectin, or a citrus pectin (all the products of Wako Pure Chemical Industry Co., Ltd.) and the obtained mixture was incubated at room temperature for 4 hours to prepare a test sample solution. As a control, 0.25 mg/ml OCIF solution (2 ml) alone was mixed with the equivalent volume of the solvent similarly and the mixture was incubated at room temperature for 4 hours. These test sample solutions were intravenously administered to 4 weeks age male Wistar rats at a dose of 2 ml/kg. After 2 and 4 hours from the administration, blood was collected from the eyehole under etherization to prepare serum. The human OCIF level in the obtained serum was measured by the ELISA described in WO96/26217. Fig. 6 shows the results.

[0049] In the same manner as above, 2 ml of 0.25 mg/ml human OCIF solution was mixed with 0.5% solution of dextran sulfate, apple pectin, or carrageenan (lambda) (all products of Wako Pure Chemical Industry Co., Ltd.) and the mixture was incubated at room temperature for 4 hours to prepare test sample solutions. As a control, 0.25 mg/ml OCIF solution alone was similarly mixed with the equivalent volume of the solvent and the mixture was incubated at room temperature for 4 hours.

These test sample solutions were intravenously administered to 4 weeks age male Wistar rats at a dose of 2 ml/kg. After 2 and 4 hours from the administration, blood was collected from the eyehole under etherization to prepare serum. The human OCIF level in the obtained serum was measured by the ELISA described in WO96/26217. Fig. 7 shows the results.

[0050] As a result, as compared with the group administered with the human OCIF solution alone, each of the groups administered with the test sample solution to which dextran sulfate, apple pectin, and citrus pectin was added, respectively, maintained evidently high circulating OCIF level (Fig. 6). The group administered with the test sample solution being added carrageenan apparently maintained high circulating OCIF level similarly (Fig. 7). Therefore, it was confirmed that the addition of the polysaccharides enables the circulating OCIF level to persist.

[Example 7]

Enhancing effect of added polysaccharides on the persistence of circulating OCIF level

[0051] Using dextran sulfate or apple pectin (Wako Pure Chemical Industry Co., Ltd.) as a polysaccharide, difference in persistence effect on circulating OCIF due to difference in administration route was studied. OCIF solutions diluted with a solvent [10 mM sodium phosphate buffer solution (pH 7.0) containing 0.15 M NaCl and 0.01% Polysorbate 80] to 50 µg/ml for intravenous administration and to 1,000 µg/ml for intramuscular administration were prepared. To these solutions the above polysaccharides were added respectively, to obtain test sample solutions. In this case, dextran sulfate was added to the OCIF solutions for intravenous and for intramuscular administration each in a concentration of 0.1%. Apple pectin was dissolved in the solvent to prepare 3 mg/ml solution and the solution was mixed with the same volume of the OCIF solution (containing 100 µg/ml and 2,000 µg/ml OCIF, respectively), for intravenous administration or intramuscular administration. The respective solutions were administered to 4 weeks age male Wistar rats at a dose of 1 ml/kg. Blood was collected from the eyehole under etherization to prepare serum after 2, 5, 10, 15, 30, 45, 60, 120, 240, and 360 minutes from the administration for the intravenous administration, and after 30 minutes, 1, 2, 3, 4, 5, 6, 8, 10, and 24 hours from the administration for the intramuscular administration. The OCIF level in the obtained serum was measured by the ELISA described in WO96/26217. Figs. 8 and 9 show the results.

[0052] As a result, in the intravenous administration (Fig. 8) and intramuscular administration (Fig. 9) of the OCIF solutions to which dextran sulfate or apple pectin was added respectively, evidently high circulating OCIF level was maintained. Therefore, it was confirmed that the addition of these polysaccharides enables the circu-

lating OCIF level to be maintained and the enhancing effect on the persistence of OCIF level is observed regardless of the administration route.

5 [Example 8]

Hypocalcemic effect of OCIF by the addition of a polysaccharide - 2

10 [0053] To 2 ml of the 0.25 mg/ml human OCIF solution prepared in the same manner as in Example 2 was mixed with an equivalent volume of 0.5% apple pectin (Wako Pure Chemical Industry Co., Ltd.) solution [solvent: 10 mM sodium phosphate buffer solution (pH 7.0) containing 0.15 M NaCl, 0.01% Polysorbate 80] and the obtained mixture was incubated at room temperature for 4 hours to prepare a test sample solution. At the same time, a mixture solution of equal volume of OCIF solution (2 ml of the 0.25 mg/ml human OCIF) and the solvent, and the solvent alone were provided. The respective test sample solutions were intravenously administered to 4 weeks age male Wistar rats at a dose of 2 ml/kg. After 4 hours from the administration, blood was collected from the eyehole under etherization to prepare serum. The OCIF level in the obtained serum was measured using Calcium C Test (Wako Pure Chemical Industry Co., Ltd.). Fig. 10 shows the results. As a result, the group administered with the 0.5 mg/kg human OCIF solution alone showed no hypocalcemic effect but in the group administered with the 0.5 mg/kg human type OCIF to which apple pectin was added, significant hypocalcemic effect was observed. Therefore, it was confirmed that the addition of the polysaccharides enhances the hypocalcemic action of human type OCIF.

INDUSTRIAL APPLICABILITY

[0054] According to the present invention, there is provided a bone-pathobolism treating agent comprising at least one substance selected from the group consisting of osteoclastogenesis inhibitory factor, its homologs, and its variants and a polysaccharide or its derivatives. According to the present invention, there is provided a therapeutic agent having excellent effect on bone-pathobolism such as osteoporosis, hypercalcemia, or chronic articular rheumatism and persistence of the activity. The therapeutic agent is useful as a medicine.

Claims

1. A bone-pathobolism treating agent comprising at least one substance selected from the group consisting of osteoclastogenesis inhibitory factor (OCIF), its homologs, and its variants and a polysaccharide or its derivatives.
2. The bone-pathobolism treating agent according to

claim 1, wherein OCIF is human OCIF.

3. The bone-pathobolism treating agent according to claim 1, wherein the polysaccharide is heparin, pectin, and/or carrageenan.

5

4. The bone-pathobolism treating agent according to claim 1, wherein the polysaccharide derivative is dextran sulfate.

10

5. A method for enhancing the activity of osteoclastogenesis inhibitory factor utilizing a polysaccharide or its derivatives.

15

20

25

30

35

40

45

50

55

Fig. 1

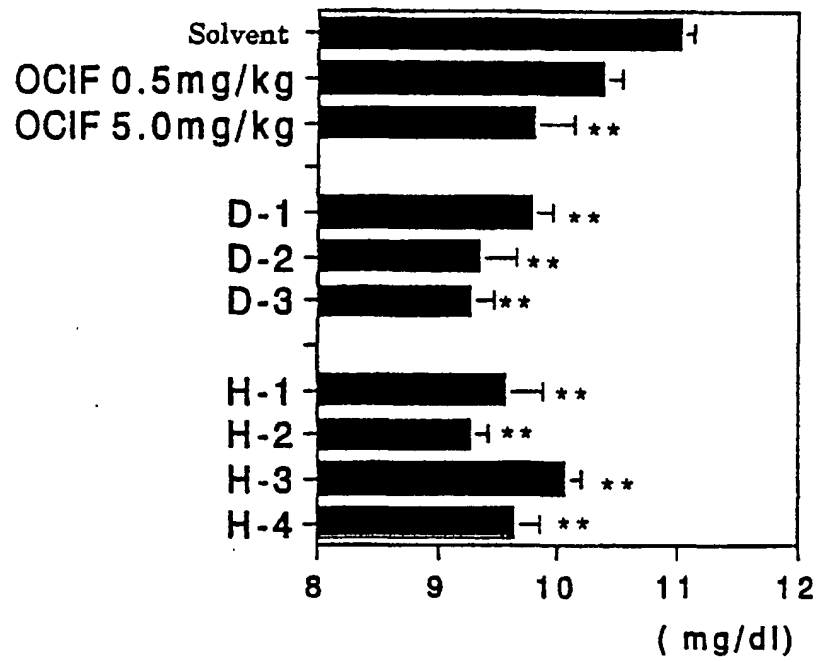


Fig. 2

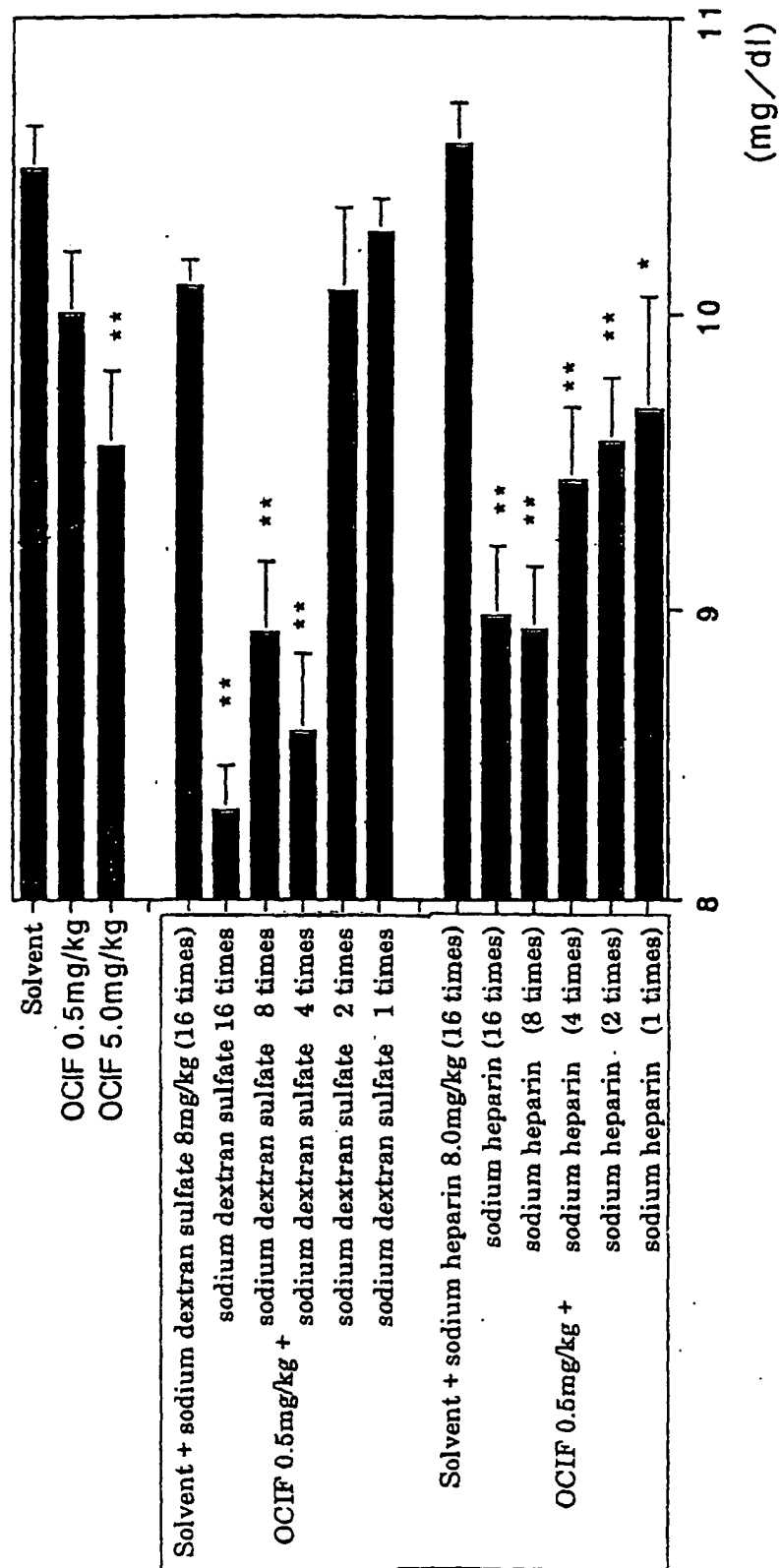


Fig. 3

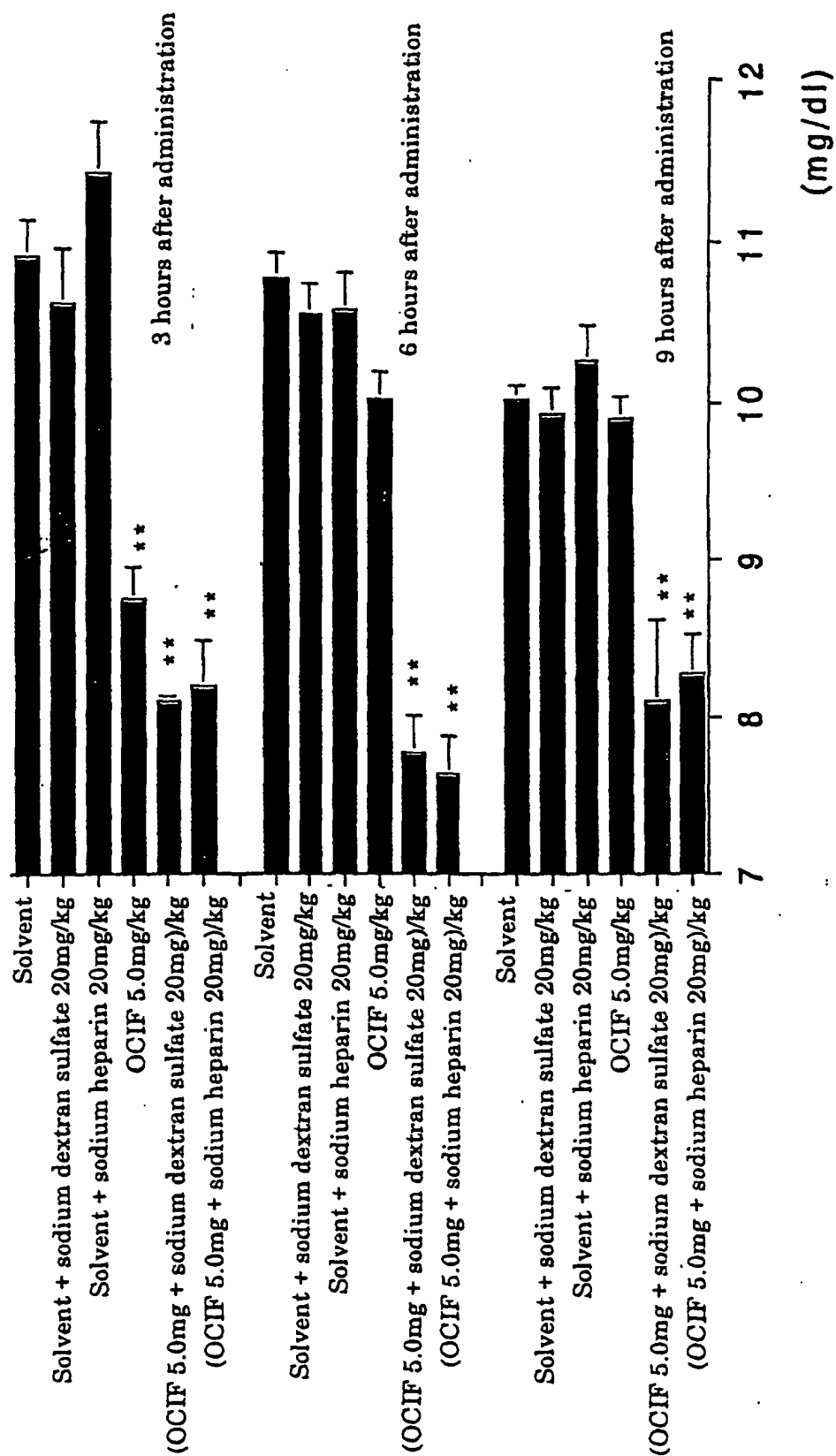


Fig. 4

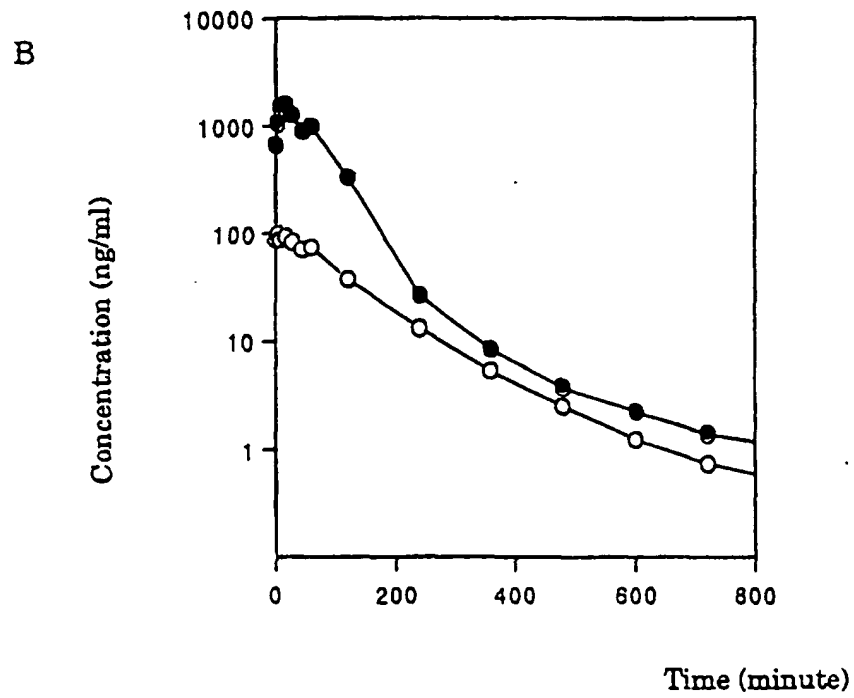
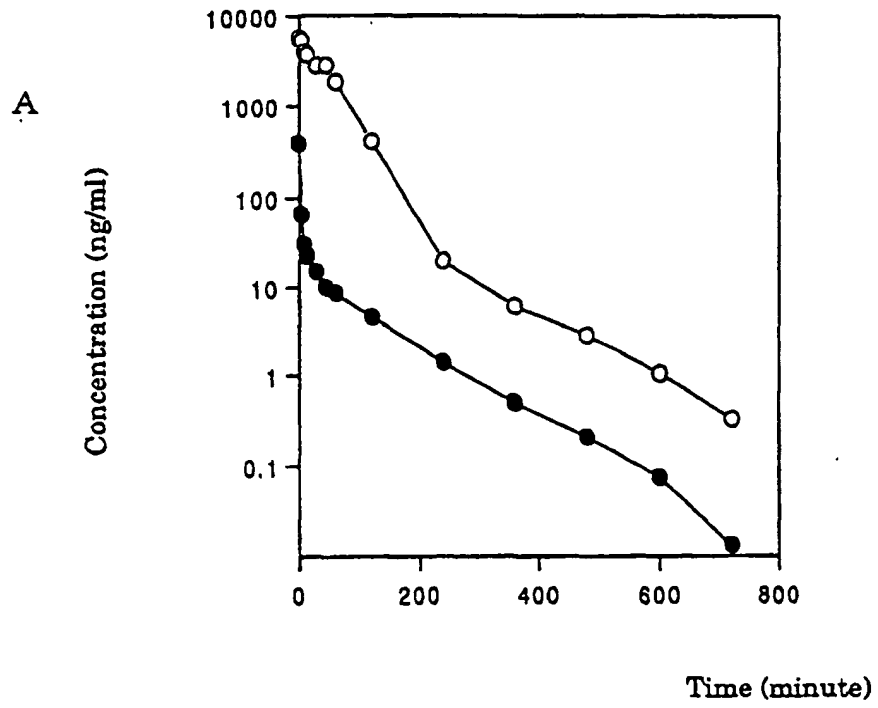


Fig. 5

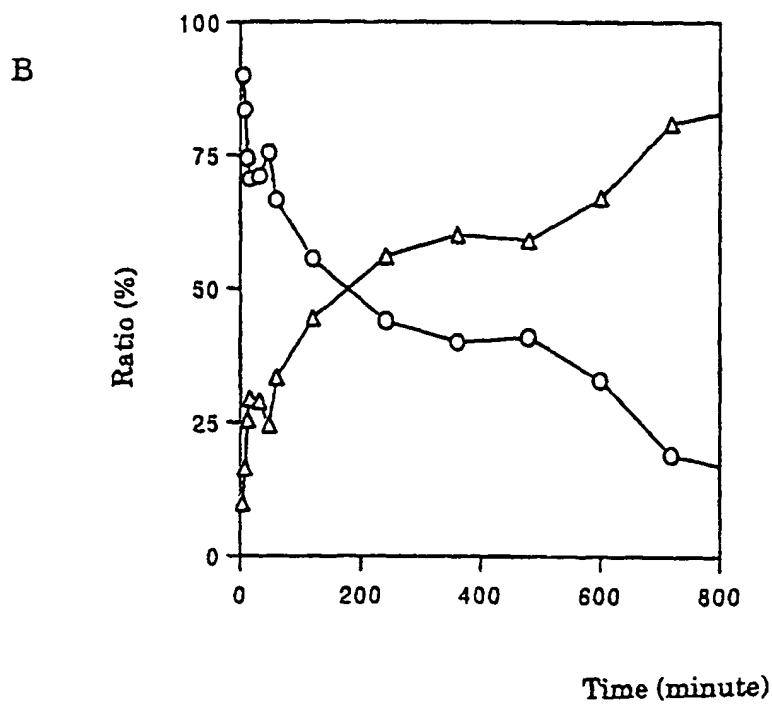
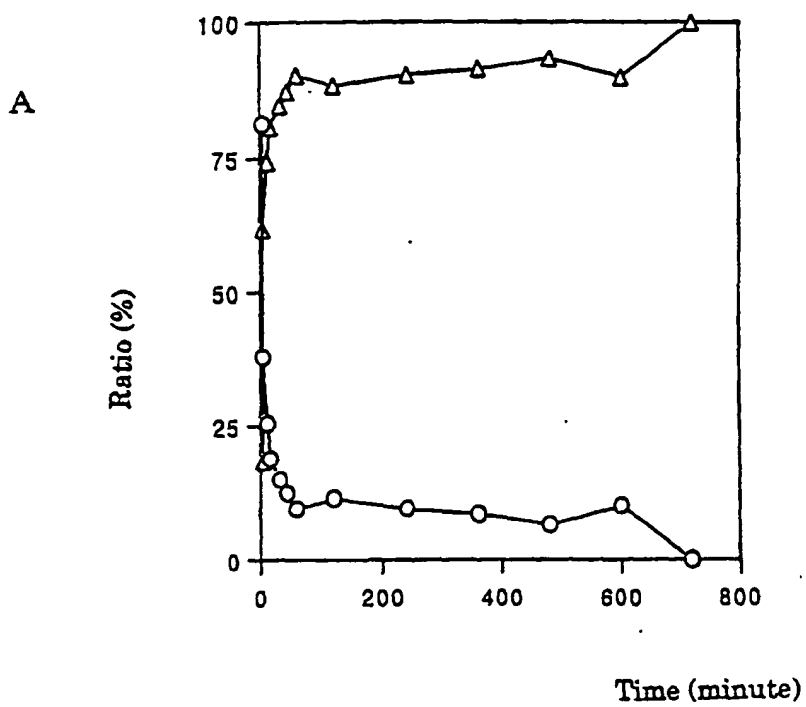


Fig. 6

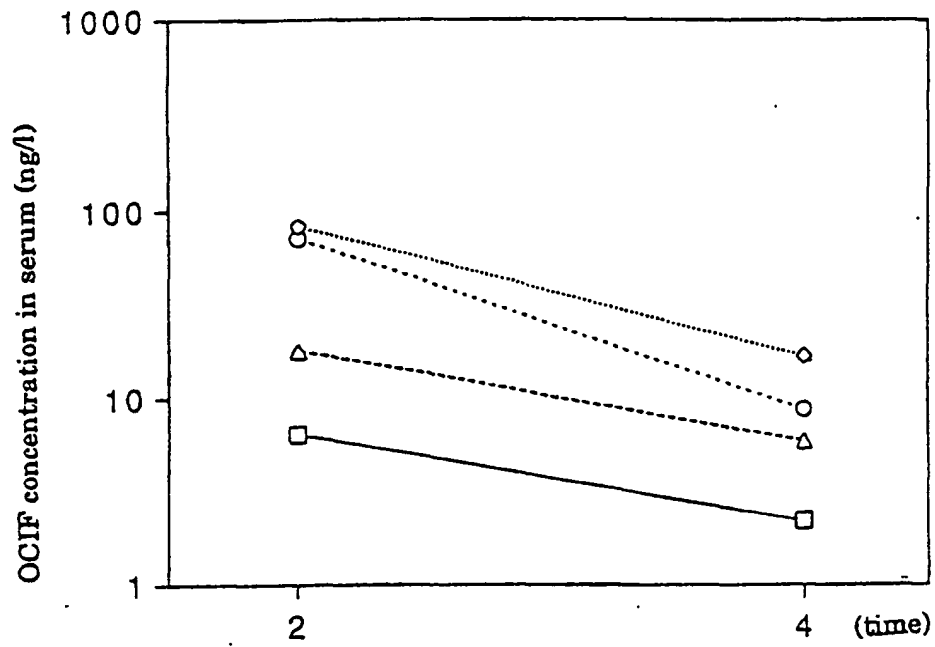


Fig. 7

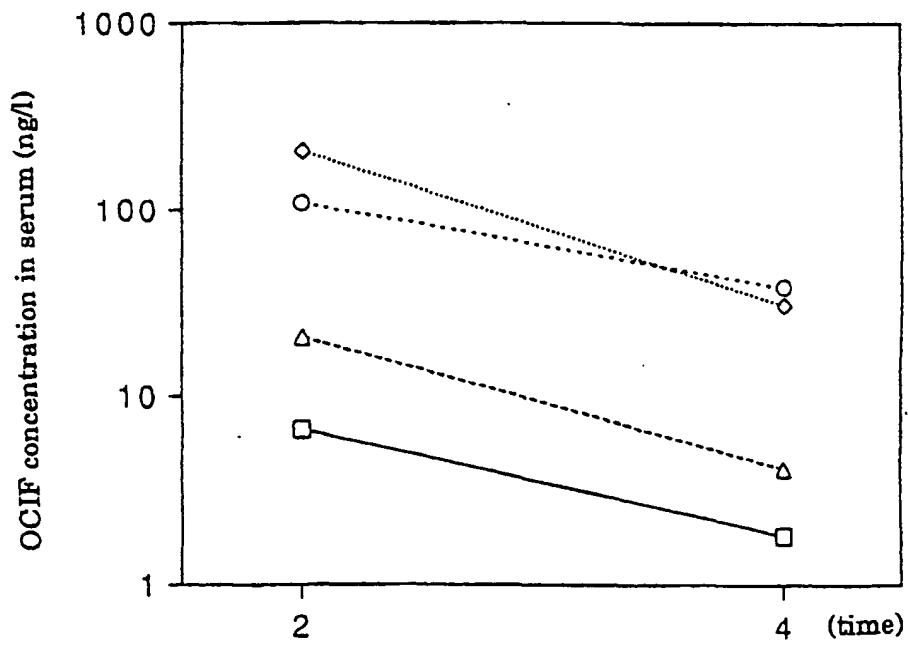


Fig. 8

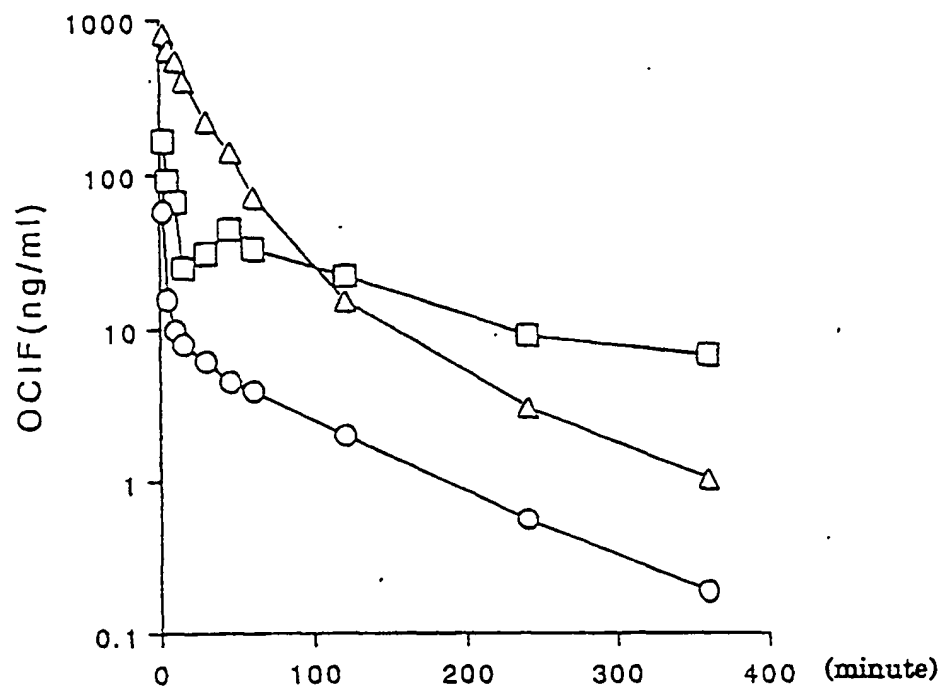


Fig. 9

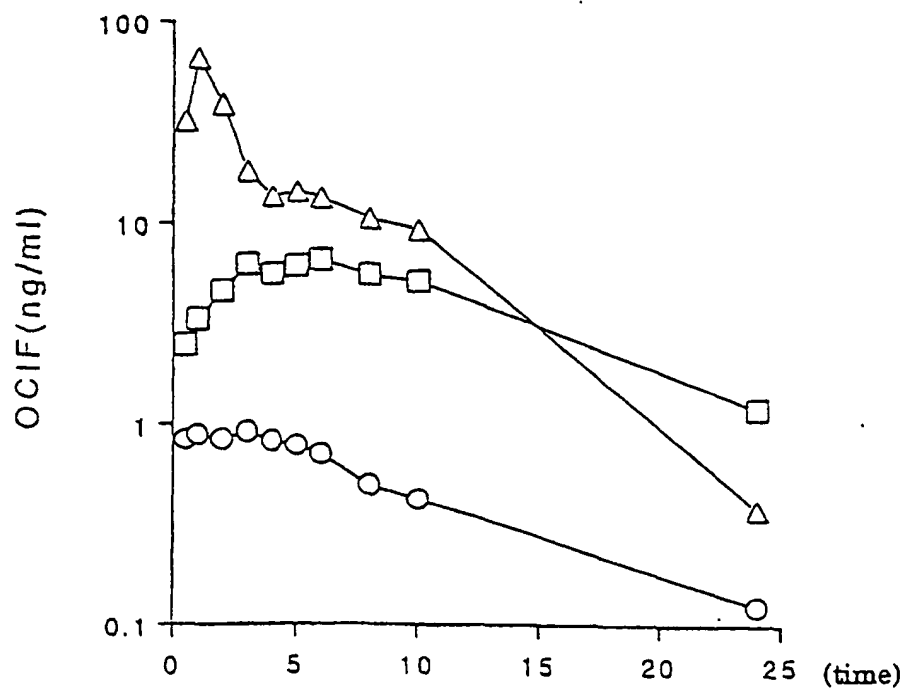
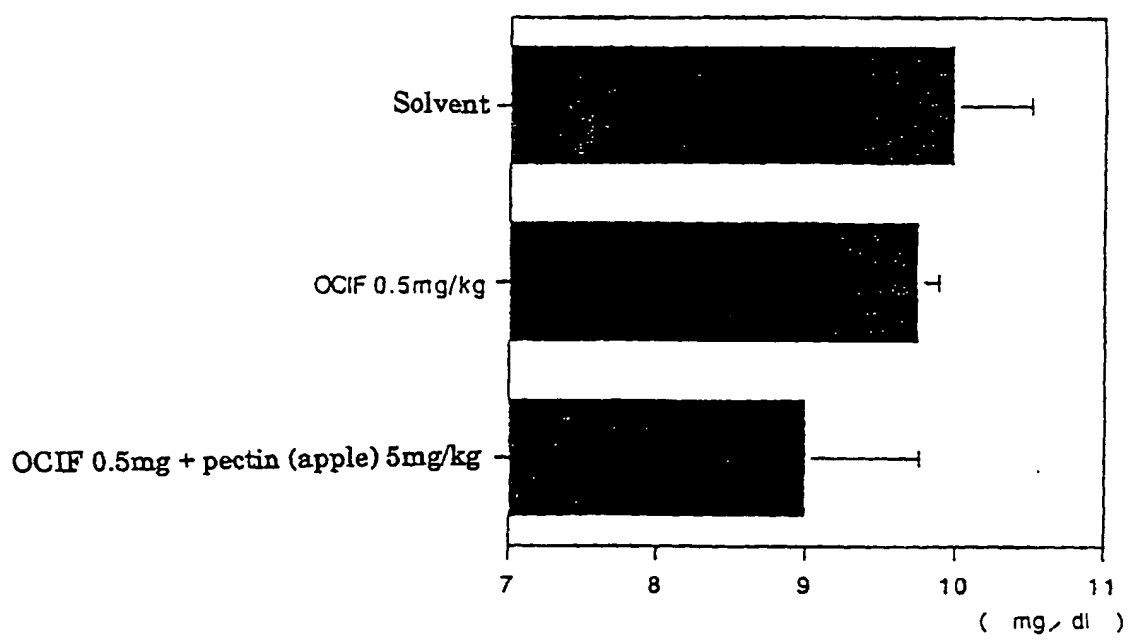


Fig. 10



INTERNATIONAL SEARCH REPORT

International application No.

PCT/JP99/05963

A. CLASSIFICATION OF SUBJECT MATTER Int.Cl. ⁷ A61K38/19, 47/36, A61P19/08, 3/00//C07K14/52		
According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) Int.Cl. ⁷ A61K38/19, 47/36, A61P19/08, 3/00//C07K14/52		
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched		
Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) CAPLUS (STN)		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	EP, 816380, A1 (SNOW BRAND MILK PRODUCTS CO., LTD), 07 January, 1998 (07.01.98) & WO, 96/26217, A1	1-4
A	WO, 98/46211, A1 (AMGEN INC.), 22 October, 1998 (22.10.98) (Family: none)	1-4
A	TOMOYASU, Akihiro et al., "Characterization of Monomeric and Homodimeric Form of Osteoclastogenesis Inhibitory Factor", BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS, April 1998, Vol. 245, No. 2, pp.382-387	1-4
A	YAMAMOTO, Michiko et al., "Hypocalcemic Effect of Osteoclastogenesis Inhibitory Factor/Osteoprotegrin in the Thyroparathyroidectomized Rat", Endocrinology, September 1998, Vol. 139, No. 9, pp.4012-4015	1-4
A	CHOWDHURY, Majeedul H. et al, effects of Heparin on Osteoclast Activity, JOURNAL OF BONE AND MINERAL RESEARCH, 1992, Vol.7, No.7, pp.771-776	1-4
<input checked="" type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.		
* Special categories of cited documents: "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art "&" document member of the same patent family		
Date of the actual completion of the international search 04 January, 2000 (04.01.00)		Date of mailing of the international search report 11 January, 2000 (11.01.00)
Name and mailing address of the ISA/ Japanese Patent Office		Authorized officer
Facsimile No.		Telephone No.

INTERNATIONAL SEARCH REPORT

International application No.

PCT/JP99/05963

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	COCHRAN, D. I. et al., "Modulation of Bone Resorption by Glycosaminoglycans: Effects of Parathyroid Hormone and Interleukin-1, Bone, 1988, Vol. 9, No. 5, pp.331-5	1-4
A	JP, 62-201825, A (LION CORPORATION), 05 September, 1987 (05.09.87) (Family: none)	1-4
A	Chem. abstr., Vol. 123, No. 18, 30 October 1995 (Columbus, OH, USA), page 662, column 2, the Abstract No. 237583p & IL, 91438, A1	1-4

INTERNATIONAL SEARCH REPORT

International application No.

PCT/JP99/05963

Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.: 5
because they relate to subject matter not required to be searched by this Authority, namely:

The subject matter of claim 5 relates to a method for treatment of the human body by therapy, which does not require an international search report by this International Search Authority in accordance with PCT Article 17(2) (a)(i) and Rule 39.1(iv).
2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest ☐ The additional search fees were accompanied by the applicant's protest.
☐ No protest accompanied the payment of additional search fees.